

Plasmid-mediated quinolone resistance among Enterobacteriaceae in Australia; Results of the AGAR Studies (2008-2010)

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INTRODUCTION

Quinolone resistance is most commonly due to mutations in DNA gyrase and topoisomerase IV. More recently plasmid-mediated quinolone resistance (PMQR) has emerged. PMQR may be due to the presence of *qnr* genes, *aac(6')-Ib-cr*, encoding for a variant aminoglycoside acetyltransferase enzyme; or *qepA*, which encodes for an efflux pump. The aim of this study was to examine a collection of *Enterobacteriaceae* from the AGAR studies for evidence of PMQR.

METHODS

Isolates

Clinically significant, non-duplicate isolates of *E. coli* (n=70), *Klebsiella* spp. (n=20) and *Enterobacter* spp. (n=10) were collected by each participating laboratory from outpatients with urinary tract infections (2008, 2010), or from patients hospitalised more than 48 hours (2009).

Susceptibility testing

MICs were determined using Vitek 2 (BioMerieux) AST-N083 (2008-2009) and AST-N149 cards. MICs were interpreted according to current CLSI M100 guidelines¹. Isolates with ciprofloxacin MIC ≥ 0.5 mg/L were referred to a central laboratory for molecular testing.

Plasmid-mediated quinolone resistance

All isolates received were screened for the presence of plasmid-mediated quinolone resistance (PMQR) genes using real-time PCR (LightCycler LC480, Roche) and published primers. *aac(6')-Ib-cr* genotyping was performed by simultaneous high resolution melting analysis of an unlabelled probe and full length amplicon².

ESBL resistance gene detection

All isolates were also screened for the presence of the *bla*_{TEM} and *bla*_{SHV} genes using previously reported oligonucleotide primers. A multiplex real-time TaqMan PCR³ was used to detect CTX-M-type genes. Strains were probed for plasmid-borne AmpC enzymes using the method described by Pérez-Pérez et al.⁴.

RESULTS

- Ciprofloxacin resistance among *E. coli* from outpatients with UTI increased from 4.9% in 2008 to 6.0% in 2010 (Figure 1, Table 1)
- Resistance was more prevalent among isolates collected from hospitalised patients in 2009 (approx. 10%).
- Over 94% of isolates with MIC ≥ 0.5 mg/L were received for molecular screening.
- PMQR genes were found in 13-14% of *E. coli* from outpatients with UTI, increasing to 23% among hospitalised patients.
- Klebsiella* spp. harboring PMQR genes were common (>60%) among hospitalised inpatients.
- Over 80% of PMQR genes detected among *E. coli* were *aac(6')-Ib-cr* variant (Figures 2 and 3)
- qnr* genes dominated among *Klebsiella* spp., with *qnrS* the dominant type.
- qepA* was only detected in four *E. coli* isolates, all from different institutions, in different states, and from all years.
- Only *qnr* genes (predominantly *qnrA*) were detected in *Enterobacter* spp., however, 30% were found in association with *aac(6')-Ib-cr* variant.
- There was a strong correlation between PMQR and ESBL production; 75% of PMQR containing isolates contained an ESBL. CTX-M types were dominant among both *E. coli* (72%) and *K. pneumoniae* (54%).

Table 1. Ciprofloxacin MICs and association with PMQR genes and ESBL production by survey year

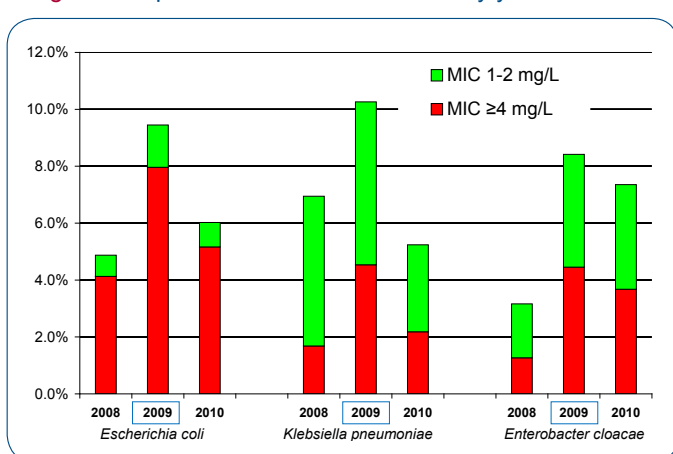
Organism	Year ^a	Percent MIC ≥ 1	PMQR ^b	Ciprofloxacin MIC (mg/L)				Total	ESBL ^c
				0.5	1	2	≥ 4		
<i>Escherichia coli</i>	2008	4.9%	-	23	9	1	66	99	16 (16%)
	(n=2,155)		+		1	1	14		16 (14%)
	2009	9.5%	-	21	19	2	97	139	42 (30%)
	(n=1746)		+	1	3	1	37		42 (23%)
	2010	6.0%	-	64	13	4	82	163	25 (15%)
	(n=2092)		+		1		24		25 (13%)
<i>Klebsiella pneumoniae</i>	2008	6.9%	-	6	13	2	3	24	6 (25%)
	(n=475)		+		6	3	5		14 (37%)
	2009	10.3%	-	2	7	2	6	17	5 (29%)
	(n=419)		+	3	7	8	12		30 (63%)
	2010	5.2%	-	13	11		9	33	6 (18%)
	(n=458)		+	2	2	1	1		6 (15%)

^a Year (number tested); 2008, 2010 community UTI; 2009, hospital inpatients

^b PMQR, Plasmid-mediated quinolone resistance gene, detected (+) or not detected (-)

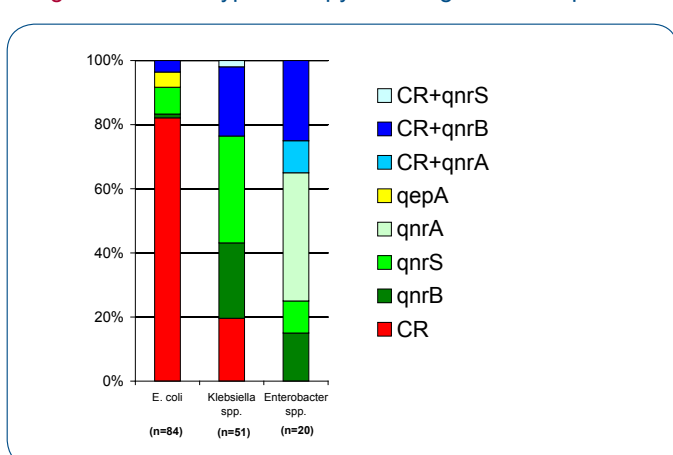
^c ESBL (TEM, SHV, CTX-M types) or plasmid-borne AmpC confirmed

Figure 2. Ciprofloxacin resistance vs study year^a



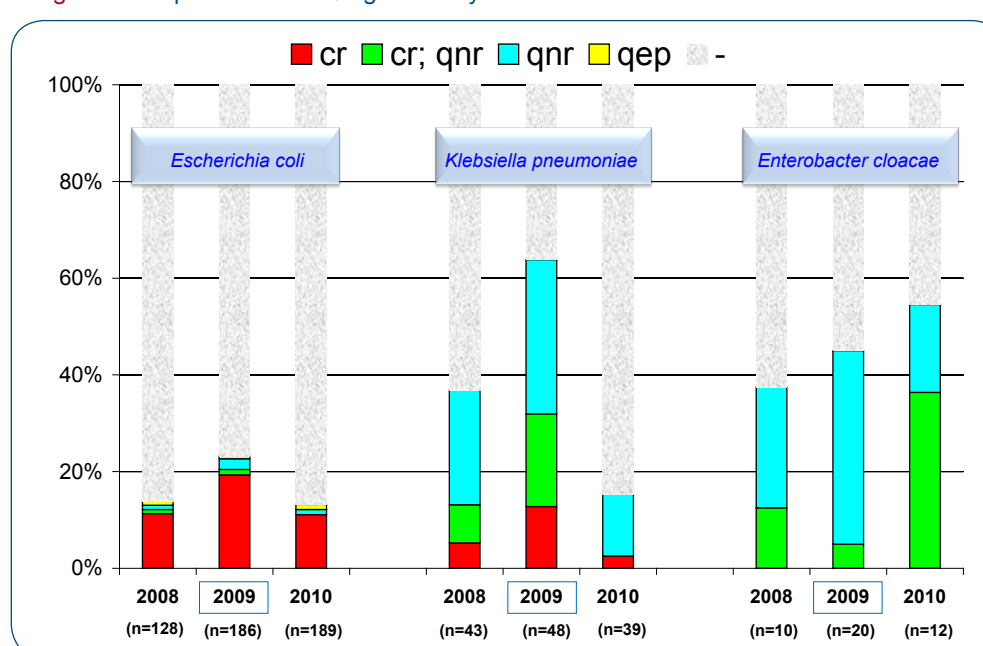
2008, 2010; outpatients with UTI; 2009; hospital inpatients

Figure 3. PMQR types/subtype among bacterial species^a



^a CR; *aac(6')-Ib-CR* variant

Figure 2. Proportion of PMQR genes vs year^a



^a 2008, 2010; outpatients with UTI; 2009; hospital inpatients (blue box). CR; *aac(6')-Ib-CR* variant

Acknowledgements

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References

- CLSI M100 S12
- Bell et al. AAC (2010) 54:1378-1380.
- Birkett et al. J Med Micro (2007) 56; 52
- Pérez-Pérez et al. J Clin Microbiol.(2002) 40:2153-2162.

CONCLUSIONS

- Although ciprofloxacin resistance is low in Australia, it is increasing
- Plasmid-mediated quinolone resistance is common, especially among hospitalised patients with *Klebsiella* spp.
- aac(6')-Ib-cr* variant is the dominant PMQR among *E. coli*
- PMQR is often associated with ESBL enzymes